A series of 3-deazapurine ribonucleosides 5a–5l bearing diverse C-substituents (alkyl, aryl and heteroaryl) in the position 6 were prepared by Pd-catalyzed cross-coupling reactions of either free 6-chloro-3-deazapurine ribonucleoside 4 or its acetyl protected congener 3 followed by deprotection. An improved synthesis of the starting 4-chloro-1-(2,3,5-tri-O-acetyl-β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (3) was developed by the application of Vorbrüggen glycosylation of silylated nucleobase with 1,2,3,5-tetra-O-acetyl-β-D-ribofuranose (2). None of compounds 5a–5l showed any considerable cytostatic or antiviral activity.

**Keywords:** Purines; Imidazo[4,5-c]pyridines; 3-Deazapurines; Nucleosides; Glycosidations; Cross-coupling reactions; Cytostatic activity.

Purine nucleosides bearing aryl or hetaryl substituents in position 6 are cytostatic. Moreover, some 6-hetarylpurine ribonucleosides also exert strong anti-HCV activities. However, the cytotoxic or cytostatic side effect prevents clinical applications as anti-HCV drugs. Therefore, in order to achieve selective inhibition of HCV RNA polymerase, some additional modifications were pursued. From the previous studies on sugar-modified derivatives it is known 2'- and 5'-deoxyribonucleosides, 3'-deoxyribo-nucleosides, as well as 2'-C-methylribonucleosides of the 6-aryl- or 6-hetarylpurine series are all inactive, while some carbocyclic homonucleosides...
were reported\textsuperscript{6} to still exert some cytostatic effects. Very recently, some L-ribonucleosides were found\textsuperscript{7} to exert weak anti-HCV effect in replicon assay but their triphosphates did not inhibit HCV RNA polymerase. Also modifications of purine ring have been pursued to show that most 2-substituted\textsuperscript{8} and 8-substituted\textsuperscript{9} 6-arylpurine ribonucleosides were inactive, while some 6-aryl-1-deazapurine nucleosides\textsuperscript{10} still exerted some activities. This shows that the N-1 nitrogen is not crucial for the interaction of these compounds with the target biological system (presumably RNA polymerase and the complementary nucleobase). Therefore the next logical step was to look into the role of N-3 nitrogen which does not make H-bonds with the complementary pyrimidine nucleobase during biosynthesis of RNA but is responsible for crucial minor groove interactions in the active site of the polymerase. In this paper we report on the synthesis and evaluation of cytostatic and anti-HCV activity of novel 6-aryl-3-deazapurine ribonucleosides. Taking into account also known cytostatic activities of 6-methylpurine\textsuperscript{11} and recently reported 6-cyclopropylpurine ribonucleosides\textsuperscript{12}, the series of 6-aryl and hetaryl derivatives was also complemented by examples of 6-alkyl-3-deazapurine nucleosides.

\begin{figure}
\centering
\includegraphics[width=\textwidth]{figure.png}
\caption{Cytostatic and anti-HCV activities of 6-aryl-3-deazapurine ribonucleosides.}
\end{figure}
3-Deazaadenosine (c3A, 4-aminoimidazo[4,5-c]pyridine) and its analogues are substrates and potent inhibitors of S-adenosyl-L-homocysteine hydrolase13 and subsequent perturbation of transmethylation reactions is at least partly responsible for their diverse biological effects. These compounds exert antiviral14, cytotoxic15, tuberculostatic16 immunosuppressive and antiinflammatory properties17. No 3-deazapurine bearing a C-substituent in position 6 was reported to the best of our knowledge.

RESULTS AND DISCUSSION

At first we had to prepare either protected or free 6-chloro-3-deazapurine riboside intermediates as starting compounds for Pd-catalyzed cross-coupling reactions with aryl(hetaryl)organometallics and boronic acids. To our surprise all the known syntheses of these nucleosides based on glycosylation of 6-chloro-3-deazapurine rely either on mercury salt method18 or fusion method19 both suffering from low overall yield, need for excess of glycosyl component, complicated separation from by-products and possible contamination by mercury salts. Therefore we were attracted by the application of the Hilbert-Johnson reaction performed under Vorbrüggen conditions20 as this approach was recently successfully employed for 3,6-difluoro-3-deazapurine21. Thus 4-chloroimidazo[4,5-c]-pyridine22 (1) was silylated by treatment with BSA in acetonitrile and then reacted with 1,2,3,5-tetra-O-acetyl-β-D-ribofuranose (2) in presence of TMSOTf at 80 °C for 1 h to afford the desired crystalline acetylated nucleoside 3 in 89% yield (Scheme 1). Deprotection of 3 by treatment with methanolic ammonia at room temperature for 24 h afforded desired free nucleoside 4 in 86% yield after crystallization.
**TABLE I**

Suzuki-Miyaura reaction of chloride 4 with boronic acids under Shaughnessy conditions

<table>
<thead>
<tr>
<th>Entry</th>
<th>R'-B(OH)₂</th>
<th>R</th>
<th>Reaction time, h</th>
<th>Cross-Coupling Product (yield)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td></td>
<td></td>
<td>3</td>
<td>5a (92%)</td>
</tr>
<tr>
<td>2</td>
<td></td>
<td></td>
<td>3</td>
<td>5b (88%)</td>
</tr>
<tr>
<td>3</td>
<td></td>
<td></td>
<td>3</td>
<td>5c (90%)</td>
</tr>
<tr>
<td>4</td>
<td></td>
<td></td>
<td>2</td>
<td>5d (72%)</td>
</tr>
<tr>
<td>5</td>
<td></td>
<td></td>
<td>2</td>
<td>5e (78%)</td>
</tr>
<tr>
<td>6</td>
<td></td>
<td></td>
<td>24</td>
<td>5f (50%)</td>
</tr>
<tr>
<td>7</td>
<td></td>
<td></td>
<td>15</td>
<td>5g (61%)</td>
</tr>
<tr>
<td>8a</td>
<td></td>
<td></td>
<td>24</td>
<td>5h (10%)&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>9</td>
<td></td>
<td></td>
<td>20</td>
<td>5i (96%)</td>
</tr>
</tbody>
</table>

<sup>a</sup> 82% of starting material 4 recovered.
With starting compounds in hands we have performed cross-coupling reactions leading to our desired 6-substituted derivatives. At first, we attempted aqueous Suzuki–Miyaura reactions of free ribonucleoside 4 under the Shaughnessy conditions. The treatment of 6-chloro-3-deazapurine riboside 4 with diverse aryl- or hetarylboronic acids in the presence of Pd(OAc)$_2$, TPPTS, and Na$_2$CO$_3$ in H$_2$O–CH$_3$CN (2:1) at 100 °C provided desired 6-aryl(hetaryl)-3-deazapurine ribosides 5a–5i (Table I). The reactions proceeded smoothly and cleanly for phenyl-, 3-pyridyl- and for 2- and 3-furyl- and thienylboronic acids, providing the desired products in high yields (entries 1–5, 9). For 2-pyrrolyl and 3-pyrrolyl the isolated yields of products 5f and 5g were only moderate (entries 6, 7), but all starting chloride 4 was always fully consumed indicating possible partial decomposition of products 5f and 5g under reaction conditions. It should be also noted, that N-protecting groups in both starting pyrrolylboronic acids were simultaneously removed under the conditions of coupling (BOC for 2-pyrrolyl and triisopropylsilyl for 3-pyrrolyl). The reaction of 4 with 1H-pyrazole-5-boronic acid was very sluggish giving product 5h (entry 8) in only 10% yield after crystallization and 82% of starting chloride 4 was recovered.

While the attempts to introduce similarly methyl and ethyl substituents by Suzuki reaction of 4 with corresponding alkylboronic acids under above mentioned conditions failed (in the case of methyl only trace of the product was obtained and for ethyl no reaction was observed) we turned to reactions with trialkylaluminums known to be suitable for introduction of alkyl substituents to purines. Thus acetylated 6-chloro-3-deazapurine riboside 3 was reacted with trimethyl- and triethylaluminums in the presence of Pd(PPh$_3$)$_4$ in refluxing THF affording 6-alkylpurine nucleosides 6a and 6b in 53 and 60% yields, respectively (Scheme 2). These acetylated products 6a and 6b were deprotected with catalytic sodium methoxide in methanol giving the desired nucleosides 5j and 5k in 71 and 70% yields, re-

**Scheme 2**

spectively, after recrystallization. For the preparation cyclopropyl derivative Negishi reaction between chloride 3 and cyclopropylzinc chloride in the presence of Pd(PPh3)4 was conducted to furnish protected cyclopropyl derivative 6c in moderate 38% yield and subsequent deprotection gave free nucleoside 5l (70%).

It should be noted, that some of the attempted cross-couplings (successful in 6-chloropurines) failed here with 6-chloro-3-deazapurines 3 or 4. The Stille reaction of acetylated nucleoside 3 with 2-(tributylstannyl)thiazole or 2-(tributylstannyl)pyridine and the Negishi reaction with benzyloxy-methylzinc iodide can serve as example. Markedly decreased reactivity of 3-deazapurine derivatives compared to corresponding purines in nucleophilic substitutions of 6-chloro group is known.

All the title 6-substituted 3-deazapurine ribonucleosides 5a–5l were subjected to biological activity screening. The cytostatic activity in vitro (inhibition of cell growth) was studied on the following cell cultures: mouse leukemia L1210 cells (ATCC CCL 219), human promyelocytic leukemia HL60 cells (ATCC CCL 240), human cervix carcinoma HeLaS3 cells (ATCC CCL 2.2) and human T lymphoblastoid CCRF-CEM cell line (ATCC CCL 119). Antiviral activity of nucleosides 5a–5l was evaluated in a HCV subgenomic replicon assay. None of the compounds showed any considerable cytostatic or antiviral activity in these assays up to 10 µM concentration. Apparently, removal of the N-3 nitrogen leads to inactive compounds indicating that some specific interactions of this nitrogen with the target biological system (most probably minor groove H-bond interaction in the active site of the RNA polymerase) are crucial for the biological activity of this class of compounds.

**EXPERIMENTAL**

NMR spectra were recorded on Bruker Avance 400 MHz (1H at 400 MHz, 13C at 100.6 MHz) and Bruker Avance 500 MHz (500 MHz for 1H and 125.7 MHz for 13C) spectrometers. Chemical shifts (in ppm, δ-scale) were referenced to TMS as internal standard. Coupling constants (J) are given in Hz. The assignment of carbons was based on C,H-HSQC and C,H-HMBC experiments. IR spectra (wavenumbers in cm⁻¹) were recorded on a Brucker IFS 88 spectrometer. Melting points were determined on a Kofler block and are uncorrected. Optical rotations were measured at 25 °C on an Autopol IV (Rudolph Research Analytical) polarimeter, [α]D values are given in 10⁻¹ deg cm² g⁻¹. High resolution mass spectra (HR MS) were measured on a LTQ Orbitrap XL (Thermo Fisher Scientific) spectrometer using electrospray ionization. High performance flash chromatography (HPFC) purifications were performed using SP1™ Flash Purification System (Biotage) on C-18 columns using water-methanol gradient. THF was dried and freshly distilled from sodium/benzophenone.
4-Chloro-1-(2,3,5-tri-O-acetyl-β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (3)

To a slurry of 4-chloro-1H-imidazo[4,5-c]pyridine (1; 1.69 g, 11 mmol) in dry CH₂CN (50 ml), BSA (2.72 ml, 11 mmol) was added and the mixture was stirred at RT for 20 min (clear solution), followed by addition of tetracetyl ribose 2 (3.85 g, 12.1 mmol). TMSOTf (1.99 ml, 11 mmol) was then added dropwise at 0 °C and the mixture was heated at 80 °C for 3 h. After cooling and dilution with CHCl₃ (100 ml) the mixture was washed with saturated aqueous NaHCO₃ (50 ml). Aqueous phase was re-extracted with CHCl₃ (2 × 10 ml), collected organics were dried over MgSO₄ and final column chromatography on silica (hexanes-AcOEt, 1:1) provided nucleoside 3 (4.05 g, 89%) as a colorless crystalline solid, m.p. 160–161 °C (hexane-AcOEt); ref.19 158.5–159 °C. 1H NMR (400 MHz, CDCl₃): 2.10, 2.15, 2.16 (3 × s, 3 × 3H, CH₃CO); 4.39 (dd, 1 H, J⁺= 6.3, J⁻= 5.3, J₂,₂' = 5.3, 5.1, 5.3, 5.1, 5.3, 4, 3, 3', 5, 5', 5), 5.11 (t, 1 H, J₂,₂' = 5.1, J₂,₂' = 5.3, 5.3, 5.3, 5.1, 5.1, 5.3, 5.1, 5.3, H-3); 5.51 (t, 1 H, J₂,₂' = 5.3, 5.3, 5.3, 5.1, 5.1, 5.3, 5.1, 5.3, H-2); 6.07 (d, 1 H, J₁,₁' = 5.3, J₁,₁' = 5.1, 5.3, 5.1, 5.1, 5.3, 5.1, 5.3, H-1); 7.53 (d, 1 H, J₁,₁' = 5.6, H-7); 8.23 (d, 1 H, J₁,₁' = 5.6, H-6); 8.27 (s, 1 H, H-2). 13C NMR (100 MHz, CDCl₃): 20.32, 20.45 and 20.73 (3 × CH₃); 54.46, 54.57 and 54.64 (3 × C₇); 61.59 (CH₂-5); 120.2, 111.4, 108.2, 104.3 (C₆); 141.55 (C-1); 141.59 (C-6); 145.07 (CH-2). HR MS (ESI): calculated for C₁₁H₇ClN₂NaO₄ [M + Na] 308.0414, found 308.0408. For C₁₁H₁₀ClN₂O₄ calculated: 46.25% C, 4.23% H, 14.71% N; found: 46.11% C, 4.26% H, 14.31% N.

4-Chloro-1-(β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (4)

Acetylated nucleoside 3 (3.29 g, 7.99 mmol) was treated with methanolic ammonia (24%, 70 ml) at RT for 24 h. After removal of volatiles under reduced pressure, the product was crystallized from 96% ethanol affording chloro riboside 4 (1.96 g, 86%) as colorless prisms, m.p. 192–193 °C; ref.19 192.5–193.5 °C. [α]D -39.1 (c 0.13, DMSO); ref.18 [α]D -39.1 (c 1.02, MeOH); ref.19 [α]D -41.6 (c 1.25, MeOH). 1H NMR (400 MHz, DMSO-d₆): 3.64 (ddd, 1 H, J⁺= 4.6, OH-3, 3, 3', 5, 5', 5); 4.12 (ddd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 4.61 (ddd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 4.91 (td, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.02 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.20 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.27 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.38 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.54 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.58 (dd, 1 H, J⁺= 3.5, 3, 3', 5, 5', 5); 5.93 (d, 1 H, J⁺= 6.3, 3, 3', 5, 5', 5); 7.51 (d, 1 H, J⁺= 5.5, 3, 3', 5, 5', 5); 8.17 (d, 1 H, J⁺= 5.5, 3, 3', 5, 5', 5); 8.70 (s, 1 H, H-2). 13C NMR (100 MHz, DMSO-d₆): 61.59 (CH₂-5); 70.63 (CH-3); 74.69 (CH-2); 86.62 (CH-4); 89.73 (CH-1); 108.25 (CH-7); 138.00 (C-3a); 138.63 (C-7a); 141.78 (CH-2); 142.08 (CH-6); 143.34 (C-4); 169.27, 169.46 and 170.00 (3 × C=O). IR (CCl₄): 2959, 2927, 2855, 1759, 1234, 1219, 1211, 967. HR MS (ESI): calculated for C₁₁H₁₀ClN₂O₄ [M + Na] 308.0414, found 308.0408. For C₁₁H₁₀ClN₂O₄ calculated: 46.25% C, 4.23% H, 14.71% N; found: 46.11% C, 4.26% H, 14.31% N.

Preparation of 6-Aryl(hetaryl)-3-deazapurine Ribosides 5a-Si

General Procedure

To an argon purged flask containing 6-chloro-3-deazapurine riboside 4 (214 mg, 0.75 mmol), boronic acid (0.94 mmol) and Na₂CO₃ (236 mg, 2.25 mmol), a pre-prepared solution of Pd(OAc)₂ (8 mg, 0.037 mmol) and TPPTS (53 mg, 0.093 mmol) in water-CH₂CN (2:1, 4 ml) was added. The reaction mixture was stirred at 100 °C for 2–24 h. After cooling the mixture...
was neutralized by the addition of aqueous HCl (3 m solution) and after concentration in vacuo final purification by reverse phase chromatography afforded products 5a-5i.

4-Phenyl-1-(β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (5a). Yield 92%. Reaction time 3 h. White solid after lyophilization, m.p. 99–104 °C. [α]D = -59.5 (c 1.6, DMSO). 1H NMR (400 MHz, DMSO-d6): 3.66 (ddd, 1 H, Jgem = 12.0, J5′b,OH = 5.1, J5′b,4′ = 3.5, H-5′b); 3.71 (ddd, 1 H, Jgem = 12.0, J5a,OH = 5.3, J5a,4′ = 3.5, H-5′a); 4.03 (td, 1 H, J4′,5 = 3.5, J4′,3′ = 3.1, H-4′); 4.16 (ddd, 1 H, J3′,2′ = 5.1, J3′,OH = 4.7, J3′,4′ = 3.1, H-3′); 4.42 (ddd, 1 H, J2′,OH = 6.4, J2′,1′ = 6.3, J2′,3′ = 5.1, H-2′); 5.21 (dd, 1 H, JOH,5′a = 5.3, JOH,5′b = 5.1, OH-5′); 5.30 (d, 1 H, JOH,3′ = 4.7, OH-3′); 5.57 (d, 1 H, JOH,2′ = 6.4, OH-2′); 5.97 (d, 1 H, J1′,2′ = 6.3, H-1′); 7.42–7.56 (m, 3 H, H-m,p-Ph); 7.82 (d, 1 H, J6′ = 5.5, H-7); 8.46 (d, 1 H, J6′ = 5.5, H-6); 6.88–6.72 (m, 3 H, H-2 and H-o-Ph). 13C NMR (100.6 MHz, DMSO-d6): 61.17 (CH-5′); 70.15 (CH-3′); 73.97 (CH-2′); 85.89 (CH-4′); 88.87 (CH-1′); 106.50 (CH-7); 128.19 (CH-m-Ph); 128.91 (CH-o-Ph); 128.97 (CH-p-Ph); 137.62 (C-i-Ph); 138.26 (C-3a); 138.95 (C-7a); 141.43 (CH-6); 143.64 (CH-2); 147.72 (C-4). IR (KBr): 1592, 1578, 1569, 1543, 1524, 1491, 1460, 1436, 1404, 1392, 1372, 1351, 1327, 1304, 1275, 1262, 1241, 1221, 1186, 1153, 1100, 980, 855, 832, 811, 798, 785, 772, 759, 746, 734, 721, 698, 685, 662, 591, 570, 550, 531, 511. HR MS (ESI): calculated for C17H18N3O4 [M + H] 328.1297, found 328.1295. For C17H17N3O4·0.7H2O calculated: 147.72 (C-4). IR (KBr): 1602, 1585, 1574, 1301, 1221, 1100, 1060. HR MS (ESI): calculated for C15H15N3NaO5 [M + Na] 340.0909, found 340.0905.
Cytostatic and Antiviral 6-Arylpurine Ribonucleosides

4-(Furan-3-yl)-1-(β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (5d). Yield 72%. Reaction time 2 h. Cotton-like white crystals from water, m.p. 181-186 ºC. [α]D -59.3 (c 1.8, DMSO).

1H NMR (400 MHz, DMSO-d6): 3.65 (ddd, 1 H, Jgem = 12.0, J5b,OH = 5.1, J5b,4 = 3.5, H-5′b); 3.7 (ddd, 1 H, Jgem = 12.0, J5a,OH = 5.3, J5a,4 = 3.5, H-5′a); 4.04 (td, 1 H, J4,5 = 3.5, J4,3 = 3.1, H-4′); 4.15 (ddd, 1 H, J5,2 = 5.1, J5,3 = 4.7, J3,4 = 3.1, H-3′); 4.39 (dd, 1 H, J2,OH = 6.4, J2,3 = 5.1, H-2′); 5.19 (dd, 1 H, JOH,5,2′ = 5.3, JOH,5b′ = 5.1, H-5′); 5.28 (d, 1 H, JOH,3′ = 4.7, H-3′); 5.55 (d, 1 H, JOH,2′ = 6.4, H-2′); 5.94 (d, 1 H, J2,3′ = 6.3, H-1′); 7.33 (dd, 1 H, J4,5 = 1.8, J4,2 = 0.8, H-4-furyl); 7.73 (d, 1 H, J5,6 = 5.5, H-7); 7.82 (dd, 1 H, J5,4 = 1.8, J5,2 = 1.6, H-5-furyl); 8.35 (d, 1 H, J6,7 = 5.6, H-6); 8.66 (s, 1 H, H-2); 8.75 (dd, 1 H, J2,5 = 1.6, J2,4 = 0.8, H-2-furyl). 13C NMR (100.6 MHz, DMSO-d6): 61.17 (CH2-5′); 70.16 (CH-3′); 74.00 (CH-2′); 85.89 (CH-4′); 88.89 (CH-1′); 105.85 (CH-7); 109.40 (CH-4-furyl); 124.62 (C-3-furyl); 137.43 (C-3a); 138.01 (C-7a); 141.47 (CH-6); 143.08 (C-4); 143.60 (2 C, CH-2′ and CH-3′).

1H NMR of 4-(Thiophen-3-yl)-1-(β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (5e). Yield 78%. Reaction time 2 h. Yellowish crystals from water, m.p. 105-109 ºC. [α]D -66.5 (c 2.6, DMSO).

1H NMR (400 MHz, DMSO-d6): 3.65 (ddd, 1 H, Jgem = 12.0, J5b,OH = 5.1, J5b,4 = 3.5, H-5′b); 3.71 (ddd, 1 H, Jgem = 12.0, J5a,OH = 5.4, J5a,4 = 3.5, H-5′a); 4.03 (td, 1 H, J4,5 = 3.5, J4,3 = 3.1, H-4′); 4.16 (ddd, 1 H, J5,2 = 5.1, J5,3 = 4.7, J3,4 = 3.1, H-3′); 4.41 (dd, 1 H, JOH,2 = 6.4, J2,1 = 6.3, J2,3 = 5.1, H-2′); 5.20 (dd, 1 H, JOH,5,2′ = 5.4, JOH,5b′ = 5.1, H-5′); 5.29 (d, 1 H, JOH,3′ = 4.7, H-3′); 5.56 (d, 1 H, JOH,2′ = 6.4, H-2′); 5.95 (d, 1 H, J2,3′ = 6.3, H-1′); 7.65 (dd, 1 H, J5,4 = 5.1, J5,5 = 3.0, H-5-thienyl); 7.76 (d, 1 H, J5,6 = 5.6, H-7); 8.21 (dd, 1 H, J4,5 = 5.1, J4,2 = 1.1, H-4-thienyl); 8.38 (d, 1 H, J6,7 = 5.6, H-6); 8.70 (s, 1 H, H-2); 8.83 (dd, 1 H, J5,3 = 3.0, J3,4 = 1.1, H-2-thienyl). 13C NMR (100.6 MHz, DMSO-d6): 61.18 (CH2-5′); 70.16 (CH-3′); 74.00 (CH-2′); 85.89 (CH-4′); 88.89 (CH-1′); 106.07 (CH-7); 125.80 (CH-5-thienyl); 126.89 (CH-3-thienyl); 127.63 (CH-4-thienyl); 137.44 (C-3a); 138.58 (C-7a); 140.01 (C-3-thienyl); 141.39 (CH-6); 143.71 (CH-2′); 144.50 (C-4). [α]D (c 1.9, CH3OH) -15°. Yield 78%. Reaction time 2 h. White crystals from MeOH, m.p. 205-206 ºC. [α]D -53.4 (c 1.9, DMSO).

1H NMR (400 MHz, DMSO-d6): 3.64 (ddd, 1 H, Jgem = 12.0, J5b,OH = 5.1, J5b,4 = 3.5, H-5′b); 3.70 (ddd, 1 H, Jgem = 12.0, J5a,OH = 5.3, J5a,4 = 3.5, H-5′a); 4.01 (td, 1 H, J4,5 = 3.5, J4,3 = 3.0, H-4′); 4.14 (ddd, 1 H, J5,2 = 5.1, J5,3 = 4.6, J3,4 = 3.0, H-3′); 4.39 (dd, 1 H, J2,OH = 6.1, J2,1 = 6.3, J2,3 = 5.1, H-2′); 5.18 (dd, 1 H, JOH,5,2′ = 5.3, JOH,5b′ = 5.1, H-5′); 5.28 (d, 1 H, JOH,3′ = 4.6, H-3′); 5.56 (d, 1 H, JOH,2′ = 6.1, OH-2′); 5.91 (d, 1 H, J1,2 = 6.3, H-1′); 6.21 (dt, 1 H, J4,3 = 3.7, J4,5 = 3.4, H-4-pyrr); 6.95 (ddd, 1 H, J5,6 = 2.7, J5,4 = 2.4, J3,5 = 1.6, H-5-pyrr); 7.38 (dd, 1 H, J3,4 = 3.7, J3,5 = 2.3, J5,3 = 1.6, H-3-pyrr); 7.58 (d, 1 H, J5,6 = 5.6, H-7); 8.26 (d, 1 H, J6,7 = 5.6, H-6); 8.63 (s, 1 H, H-2); 11.54 (br s, 1 H, NH-pyrr). 13C NMR (100.6 MHz, DMSO-d6): 61.18 (CH2-5′); 70.14 (CH-3′); 73.97 (CH-2′); 85.81 (CH-4′); 88.80 (CH-1′); 104.27 (CH-7); 109.28 (CH-4-pyrr); 111.56 (CH-5-pyrr); 120.89 (CH-5-pyrr); 129.58 (C-2-pyrr); 135.53 (C-3a); 138.17 (C-7a); 141.17 (CH-6); 142.71 (C-4); 143.08 (CH-2). [α]D (c 1.9, CH3OH) -15°. Yield 72%. Reaction time 2 h. Cotton-like white crystals from water, m.p. 181-186 ºC. [α]D -59.3 (c 1.8, DMSO).
C\textsubscript{14}H\textsubscript{15}N\textsubscript{5}O\textsubscript{4}·1.35H\textsubscript{2}O calculated: 49.22\% C, 5.22\% H, 20.50\% N; found: 49.47\% C, 4.74\% H, 16.04\% N.

- For C\textsubscript{14}H\textsubscript{15}N\textsubscript{5}O\textsubscript{4}·1.35H\textsubscript{2}O calculated: 49.22\% C, 5.22\% H, 20.50\% N; found: 49.47\% C, 4.74\% H, 16.04\% N.

- 4-(Pyrrrol-3-yl)-1-(β-o-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (5g). Yield 61\%. Reaction time 15 h. Greenish solid from water, m.p. 119–126 °C. [α\textsubscript{D}\textsubscript{2} = –48.8 (c 1.7, DMSO). ^1H NMR (400 MHz, DMSO-d\textsubscript{6}): 3.64 (dd, 1 H, J\textsubscript{gem} = 11.9, J\textsubscript{SO,H} = 5.1, J\textsubscript{SH,4} = 3.65, H-5\textsubscript{b}); 3.70 (ddd, 1 H, J\textsubscript{gem} = 11.9, J\textsubscript{SO,H} = 5.3, J\textsubscript{SH,4} = 3.6, H-5\textsubscript{a}); 4.00 (td, 1 H, J\textsubscript{4,5,6} = 3.6, J\textsubscript{3,4} = 3.2, H-4'); 4.14 (ddd, 1 H, J\textsubscript{3,2} = 5.1, J\textsubscript{3,SH} = 4.7, J\textsubscript{3,4} = 3.2, H-3'); 4.39 (ddd, 1 H, J\textsubscript{2,SH} = 6.5, J\textsubscript{2,1} = 6.3, J\textsubscript{2,3} = 5.1, H-2'); 5.17 (dd, 1 H, J\textsubscript{OH,5} = 5.3, J\textsubscript{OH,3} = 5.1, OH-5'); 5.26 (d, 1 H, J\textsubscript{OH,3} = 4.7, OH-3'); 5.54 (d, 1 H, J\textsubscript{OH,2} = 6.5, OH-2'); 5.89 (d, 1 H, J\textsubscript{1,2} = 6.3, H-1'); 6.84 (td, 1 H, J\textsubscript{5,4} = 2.6, J\textsubscript{5,2} = 2.0, H-5-pyrr); 7.03 (td, 1 H, J\textsubscript{4,5} = 2.6, J\textsubscript{4,2} = 1.5, H-4-pyrr); 7.51 (d, 1 H, J\textsubscript{6,7} = 5.6, H-7'); 8.03 (ddd, 1 H, J\textsubscript{2,3} = 2.8, J\textsubscript{2,4} = 2.0, J\textsubscript{2,5} = 1.5, H-2-pyrr); 8.23 (d, 1 H, J\textsubscript{6,7} = 5.6, H-6); 8.55 (s, 1 H, H-2); 11.10 (br s, 1 H, NH-pyrr). ^13C NMR (100.6 MHz, DMSO-d\textsubscript{6}): 61.23 (CH\textsubscript{2}-\textsubscript{5}); 70.16 (CH\textsubscript{3}-\textsubscript{3}); 73.90 (CH\textsubscript{2}-\textsubscript{5}); 85.72 (CH\textsubscript{2}-\textsubscript{4}); 88.72 (CH-1'); 103.68 (CH-7); 107.69 (CH-4-pyrr); 118.23 (CH-5-pyrr); 120.58 (CH-2-pyrr); 122.08 (C-3-pyrr); 136.47 (C-3a); 137.94 (C-7a); 141.26 (CH-6); 142.47 (CH-2); 146.96 (CH-4-pyrazolyl); 151.50 (C-3-pyrr); 153.81 (C-7-pyrr); 186.85 (CH\textsubscript{2}-\textsubscript{5}).

- 1H NMR (400 MHz, DMSO-d\textsubscript{6} + DCl): 60.97 (CH\textsubscript{2}-\textsubscript{5}); 85.99 (CH-4-pyrazolyl); 8.13 (d, 1 H, J\textsubscript{4,3} = 3.6, J\textsubscript{4,5} = 3.2, H-4'); 4.17 (dd, 1 H, J\textsubscript{3,2} = 5.1, J\textsubscript{3,4} = 3.6, H-3'); 4.37 (dd, 1 H, J\textsubscript{2,1} = 5.7, J\textsubscript{2,3} = 5.1, H-2'); 6.14 (d, 1 H, J\textsubscript{1,2} = 5.7, H-1'); 7.64 (d, 1 H, J\textsubscript{4,5} = 2.4, H-4-pyrazolyl); 8.13 (d, 1 H, J\textsubscript{5,4} = 2.4, H-5-pyrazolyl); 8.38 (d, 1 H, J\textsubscript{6,7} = 6.7, H-7'); 8.48 (d, 1 H, J\textsubscript{6,7} = 6.7, H-6); 9.17 (s, 1 H, H-2). ^13C NMR (125.7 MHz, DMSO-d\textsubscript{6} + DCl): 60.97 (CH\textsubscript{2}-\textsubscript{5}); 70.25 (CH-3'); 75.19 (CH-2'); 86.85 (CH\textsubscript{2}-\textsubscript{4}); 90.20 (CH-1'); 108.95 (CH-7); 109.25 (CH-4-pyrazolyl); 132.09 (CH-5-pyrazolyl); 134.12 (CH-6); 137.18 (C-3a); 138.05 (C-4); 140.54 (C-3-pyrazolyl); 143.30 (C-7a); 148.84 (CH-2)' IR (KBr): 3388, 1588, 1552, 1480, 1460, 1217, 1103, 1041. HR MS (ESI): calculated for C\textsubscript{15}H\textsubscript{17}N\textsubscript{4}O\textsubscript{4} [M + H]\textsubscript{+} 317.1250, found 317.1245. For C\textsubscript{15}H\textsubscript{17}N\textsubscript{4}O\textsubscript{4}·0.5H\textsubscript{2}O calculated: 55.38\% C, 5.27\% H, 17.22\% N; found: 55.49\% C, 4.92\% H, 16.95\% N.
An argon purged flask containing a mixture of acetylated 6-chloro-3-deazapurine riboside 3 (300 mg, 0.73 mmol), trimethylaluminium (2 mL solution in toluene; 0.73 mmol, 1.46 mmol) and Pd(PPh3)4 (42 mg, 0.036 mmol) in THF (4 mL) was stirred at 100 °C for 3 h. The mixture was diluted with CHCl3 (2 × 10 ml). Collected organic extracts were dried over MgSO4, volatiles were removed in vacuo and the residue was chromatographed on silica gel (AcOEt) affording product 6a as colorless oil (151 mg, 53%). 1H NMR (400 MHz, CDCl3): 2.09, 2.17 and 2.19 (3 × s, 3 × CH3); 2.90 (s, 3 H, CH3); 4.41 (dd, 1 H, Jgem = 12.6, J5b,6a = 2.8, H-5′a); 4.46 (dd, 1 H, Jgem = 12.6, J5a,5b = 3.0, H-5′a); 4.50 (ddd, 1 H, J4a,5 = 4.4, J4a,5′ = 3.0, 2.8, H-4′a); 5.43 (dd, 1 H, J2,3′ = 5.6, J3,4′ = 4.4, H-3′); 5.56 (t, 1 H, J2′,3′ = J2,3′ = 5.6, H-2′); 6.08 (d, 1 H, J1′,2 = 5.6, H-1′); 7.39 (d, 1 H, J13,6 = 5.7, H-7); 8.17 (s, 1 H, H-2); 8.35 (d, 1 H, J6,7 = 5.7, H-6). 13C NMR (100.6 MHz, CDCl3): 13.22 (CH3H3CO); 62.73 (CH2-5′); 73.29 (CH-2′); 70.11 (CH-3′); 80.37 (CH-4′a); 87.05 (CH-1′a); 140.39 (CH-2′); 142.20 (CH-6′). IR (CCl4): 1757, 1600, 1588, 1371, 1220, 1103, 1063, 1048. HR MS (ESI): calculated for C16H16N4O4·1.8H2O: 329.1250, found 329.1247. For C16H16N4O4·1.8H2O calculated: 53.27% C, 5.48% H, 15.53% N; found: 53.42% C, 5.37% H, 15.34% N.

4-Ethyl-1-(2,3,5-tri-O-acetyl-β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (6b)

An argon purged flask containing a mixture of acetylated 6-chloro-3-deazapurine riboside 3 (288 mg, 0.70 mmol), trimethylaluminium (1 mL solution in hexane; 1.4 mL, 1.4 mmol) and Pd(PPh3)4 (42 mg, 0.035 mmol) in THF (4 mL) was stirred at 100 °C for 3 h. The mixture was diluted with CHCl3 (20 ml) and treated with saturated aqueous NH4Cl (20 ml). The slurry was filtered through cellite and after phase separation, aqueous phase was re-extracted with CHCl3 (2 × 10 ml). Collected organic extracts were dried over MgSO4, volatiles were removed in vacuo and the residue was chromatographed on silica gel (AcOEt) affording product 6b as colorless oil (169 mg, 60%). 1H NMR (400 MHz, CDCl3): 1.44 (t, 3 H, Jgem = 7.6, CH3(CH2)); 2.10, 2.17, 2.18 (3 × s, 3 × H, 3 × CH3CO); 3.28 (q, 2 H, Jgem = 7.6, CH2CH3); 4.41 (dd, 1 H, Jgem = 12.5, J5b,6a = 2.8, H-5′a); 4.46 (dd, 1 H, Jgem = 12.5, J5a,5b = 3.0, H-5′a); 4.50 (ddd, 1 H, J4a,5 = 4.4, J4a,5′ = 3.0, 2.8, H-4′a); 5.43 (dd, 1 H, J2,3′ = 5.6, J3,4′ = 4.3, H-3′); 5.56 (t, 1 H, J2′,3′ = J2,3′ = 5.6, H-2′); 6.09 (d, 1 H, J1′,2 = 5.6, H-1′); 7.39 (d, 1 H, J6,7 = 5.7, H-7); 8.16 (s, 1 H, H-2); 8.39 (d, 1 H, J6,7 = 5.7, H-6). 13C NMR (100.6 MHz, CDCl3): 13.22 (CH3H3CO); 20.33, 20.50 and 20.76 (3 × CH3CO); 26.85 (CH2CH3); 62.73 (CH5); 70.11 (CH-3′); 73.30 (CH-2′); 80.37 (CH-1′a); 87.04 (CH-1′); 104.01 (CH-7); 136.92 (C-7a); 139.82 (C-3a); 140.39 (CH-2′); 142.20 (CH-6′); 152.68 (C-4); 169.20, 169.49 and 170.09 (3 × CO). IR (CCl4): 1758, 1602, 1590, 1371, 1219, 1100, 1063, 1048. HR MS (ESI): calculated for C19H24N3O7·1.8H2O: 406.1614, found 406.1609.

4-Cyclopropyl-1-(2,3,5-tri-O-acetyl-β-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (6c)

Tetrahydrofuran (4 ml) was added to flame-vacuum dried zinc chloride (340 mg, 2.5 mmol) under argon. The mixture was stirred at -10 °C and a solution of cyclopropylmagnesium
bromide (1 M solution in THF; 2.5 ml) was added dropwise. The mixture was stirred for 40 min and then a solution of a 6-chloro-3-deazapurine riboside 3 (205 mg, 0.5 mmol) and Pd(PPh₃)₄ (40 mg, 0.035 mmol) in THF (5 ml) was added. The resulting mixture was stirred at 70 °C for 8 h. Then the reaction mixture was diluted with water (50 ml) and washed with ethyl acetate (3 × 50 ml). The collected organic layers were washed with brine, dried over MgSO₄ and the residue was purified by column chromatography on silica gel (AcOEt–hexane 0–30%) providing compound 6c as a white foam (80 mg, 38%). ¹H NMR (500 MHz, DMSO-d₆): 6.4, J₅,4 = 6.4, H-5; 6.25 (d, 1 H, J₅,4 = 5.0, H-5) and then a solution of a 6-chloro-3-deazapurine riboside

Compound 6a (129 mg, 0.33 mmol) was treated with 1 M NaOMe/MeOH (100 µl, 0.1 mmol) in MeOH (2 ml) at RT for 1 h and, after removal of volatiles, the crude product was desalted by reverse phase chromatography and crystallized from water affording nucleoside 5j as colorless crystals (62 mg, 71%), m.p. 245–249 °C. [α]₉D = 68.3 (c 2.2, DMSO). ¹H NMR (400 MHz, DMSO-d₆): 2.70 (s, 3 H, CH₃); 3.62 (td, 1 H, J₅,4 = 11.9, J₅,6 = 3.5, H-5b); 3.67 (ddd, 1 H, J₅,4 = 11.9, J₆,7 = 5.3, J₅,6 = 3.5, H-5a); 3.99 (td, 1 H, J₅,4 = 3.5, J₆,7 = 3.0, H-4); 4.12 (ddd, 1 H, J₅,4 = 5.1, J₆,7 = 3.0, J₅,6 = 3.0, H-3); 4.36 (ddd, 1 H, J₅,4 = 3.0, J₆,7 = 3.0, J₅,6 = 3.0, H-2); 5.16 (dd, 1 H, J₅,4 = 5.1, J₆,7 = 3.0, H-1); 7.64 (d, 1 H, J₅,4 = 5.1, H-7); 8.19 (d, 1 H, J₅,4 = 5.1, H-6); 8.53 (s, 1 H, H-2). ¹³C NMR (100.6 MHz, DMSO-d₆): 19.53 (CH₃); 61.20 (CH₂-5); 70.15 (CH₂-5); 73.86 (CH₂-5); 88.77 (CH-1); 105.29 (CH-7); 137.02 (CH-7); 139.35 (C-3a); 140.96 (CH-6); 142.80 (CH-2); 150.65 (C-4). IR (CHCl₃) 591, 1372, 1232, 1096, 1067. HR MS (ESI): calculated for C₁₂H₁₅N₃O₄ [M + H] 418.1609, found 418.1602.

4-Methyl-1-([β-d-ribofuranosyl]-1H-imidazo[4,5-c]pyridine (5j)

Compound 6a (129 mg, 0.33 mmol) was treated with 1 M NaOMe/MeOH (100 µl, 0.1 mmol) in MeOH (2 ml) at RT for 1 h and, after removal of volatiles, the crude product was desalted by reverse phase chromatography and crystallized from water affording nucleoside 5k as colorless crystals (72 mg, 70%), m.p. 94 °C. [α]₀D = -41.0 (c 1.4, DMSO). ¹H NMR (400 MHz, DMSO-d₆): 1.31 (t, 3 H, J₅,4 = 7.6, CH₃CH₂); 3.10 (q, 2 H, J₅,4 = 7.6, CH₂CH₂); 3.62 (ddd, 1 H, J₅,4 = 11.9, J₆,7 = 3.5, J₅,6 = 3.5, H-5b); 3.67 (ddd, 1 H, J₅,4 = 11.9, J₆,7 = 3.5, J₅,6 = 3.5, J₅,6 = 3.5, H-5a); 3.99 (td, 1 H, J₅,4 = 3.5, J₆,7 = 3.5, J₅,6 = 3.0, H-4); 4.12 (ddd, 1 H, J₅,4 = 3.5, J₆,7 = 3.0, J₅,6 = 3.0, H-3); 4.36 (ddd, 1 H, J₅,4 = 3.0, J₆,7 = 3.0, J₅,6 = 3.0, H-2); 5.16 (dd, 1 H, J₅,4 = 3.0, J₆,7 = 3.0, J₅,6 = 3.0, H-1); 6.4, J₅,4 = 6.4, H-2; 6.25 (d, 1 H, J₅,4 = 5.0, H-5) and then a solution of a 6-chloro-3-deazapurine riboside

Compound 6b (149 mg, 0.37 mmol) was treated with 1 M NaOMe/MeOH (110 µl, 0.11 mmol) in MeOH (2 ml) at RT for 1 h and, after removal of volatiles, the crude product was desalted by reverse phase chromatography and crystallized from water affording nucleoside 5k as colorless crystals (72 mg, 70%), m.p. 94 °C. [α]₀D = -41.0 (c 1.4, DMSO). ¹H NMR (400 MHz, DMSO-d₆): 1.31 (t, 3 H, J₅,4 = 7.6, CH₂CH₂); 3.10 (q, 2 H, J₅,4 = 7.6, CH₂CH₂); 3.62 (ddd, 1 H, J₅,4 = 11.9, J₆,7 = 3.5, J₅,6 = 3.5, H-5b); 3.67 (ddd, 1 H, J₅,4 = 11.9, J₆,7 = 3.5, J₅,6 = 3.5, J₅,6 = 3.5, H-5a); 3.99 (td, 1 H, J₅,4 = 3.5, J₆,7 = 3.5, J₅,6 = 3.0, H-4); 4.12 (ddd, 1 H, J₅,4 = 3.5, J₆,7 = 3.0, J₅,6 = 3.0, H-3); 4.36 (ddd, 1 H, J₅,4 = 3.0, J₆,7 = 3.0, J₅,6 = 3.0, H-2); 5.16 (dd, 1 H, J₅,4 = 3.0, J₆,7 = 3.0, J₅,6 = 3.0, H-1); 6.4, J₅,4 = 6.4, H-2; 6.25 (d, 1 H, J₅,4 = 5.0, H-5) and then a solution of a 6-chloro-3-deazapurine riboside


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OH-5'; 5.26 (d, 1 H, $J_{OH,3'} = 4.6$, OH-3'); 5.51 (d, 1 H, $J_{OH,2'} = 6.5$, OH-2'); 5.88 (d, 1 H, $J_{1',2'} = 5.7$, H-1'); 7.64 (d, 1 H, $J_{7,6} = 5.7$, H-7); 8.22 (d, 1 H, $J_{6,7} = 5.7$, H-6); 8.52 (s, 1 H, H-2). 13C NMR (100.6 MHz, DMSO-d$_6$): 13.13 (CH$_3$CH$_2$); 26.11 (CH$_2$; 61.23 (CH$_2$-5'); 70.19 (CH-3'); 73.84 (CH-2'); 85.81 (CH-4'); 88.75 (CH-1'); 105.32 (CH-7); 137.21 (C-7a); 138.73 (C-3a); 141.03 (CH-6); 142.81 (CH-2); 155.40 (C-4). IR (KBr): 3280, 1605, 1593, 1493, 1477, 1415, 1312, 1217, 1119, 1087, 998. HR MS (ESI): calculated for C$_{13}$H$_{18}$N$_3$O$_4$ [M + H] 280.1297, found 280.1291. For C$_{13}$H$_{17}$N$_3$O$_4$·H$_2$O calculated: 52.52% C, 6.44% H, 14.13% N; found: 52.57% C, 6.46% H, 14.09% N.

4-Cyclopropyl-1-($\beta$-D-ribofuranosyl)-1H-imidazo[4,5-c]pyridine (5l) Compound 6c (72 mg, 0.17 mmol) was treated with 1M NaOMe/MeOH (52 µl, 0.52 mmol) in MeOH (2 ml) at RT for 1 h and, after removal of volatiles, the crude product was desalted by reverse phase chromatography and crystallized from water–MeOH affording nucleoside 5l as colorless crystals (35 mg, 70%), m.p. 107–113 °C. [α]$_D$ –43.0 (c 0.23, MeOH). 1H NMR (500 MHz, DMSO-d$_6$): 1.13 and 1.22 (2 × m, 2×2H, H-2,3-cycloprop); 2.86 (tt, 1 H, $J_{vic} = 8.1, 4.8$, H-1-cycloprop); 3.72 and 3.77 (2 × dd, 2 H, $J_{gem} = 12.0, J_{5',4'} = 3.6$, H-5'); 4.08 (td, 1 H, $J_{4',5'} = 3.6, J_{4',3'} = 3.1$, H-4'); 4.21 (dd, 1 H, $J_{1',2'} = 5.2$, J$_{3',2'} = 3.1$, H-3'); 4.44 (dd, 1 H, $J_{1',2'} = 6.4$, J$_{2',3'} = 5.2$, H-2'); 5.96 (d, 1 H, $J_{1',2'} = 6.4$, H-1'); 7.63 (d, 1 H, $J_{7,6} = 5.6$, H-7); 8.24 (d, 1 H, $J_{6,7} = 5.6$, H-6); 8.62 (s, 1 H, H-2). 13C NMR (125.7 MHz, DMSO-d$_6$): 9.84 and 9.86 (CH$_2$-2,3-cycloprop); 12.68 (CH-1-cycloprop); 61.44 (CH$_2$-5'); 70.37 (CH-3'); 74.14 (CH-2'); 86.01 (CH-4'); 89.02 (CH-1'); 104.51 (CH-7); 137.07 (C-7a); 139.23 (C-3a); 141.47 (CH-6); 143.08 (CH-2'); 155.14 (C-4'). IR (KBr): 3480, 3403, 1601, 1593, 1473, 1455, 1388, 1366, 1221, 1102, 1063, 989, 976. HR MS (ESI) calculated for C$_{14}$H$_{18}$N$_3$O$_4$ [M + H] 292.1292, found 292.1291. For C$_{14}$H$_{17}$N$_3$O$_4$·1.15H$_2$O calculated: 53.89% C, 6.23% H, 13.47% N; found: 53.83% C, 6.22% H, 13.43% N.

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REFERENCES


